

Genomic surveillance of antimicrobial resistance in *Shigella sonnei* in Belgium 2013-2019

Natalie Fischer, PhD

EUPHEM Fellow, Cohort 2019

Sciensano, Belgium



Sciensano National Reference Laboratory for *Salmonella* and *Shigella*
Sciensano Service of Epidemiology of Infectious Diseases

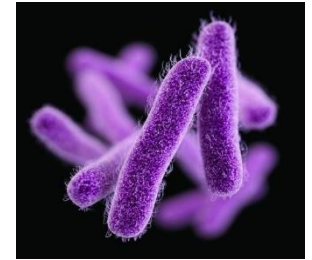
.be

Objectives of this webinar

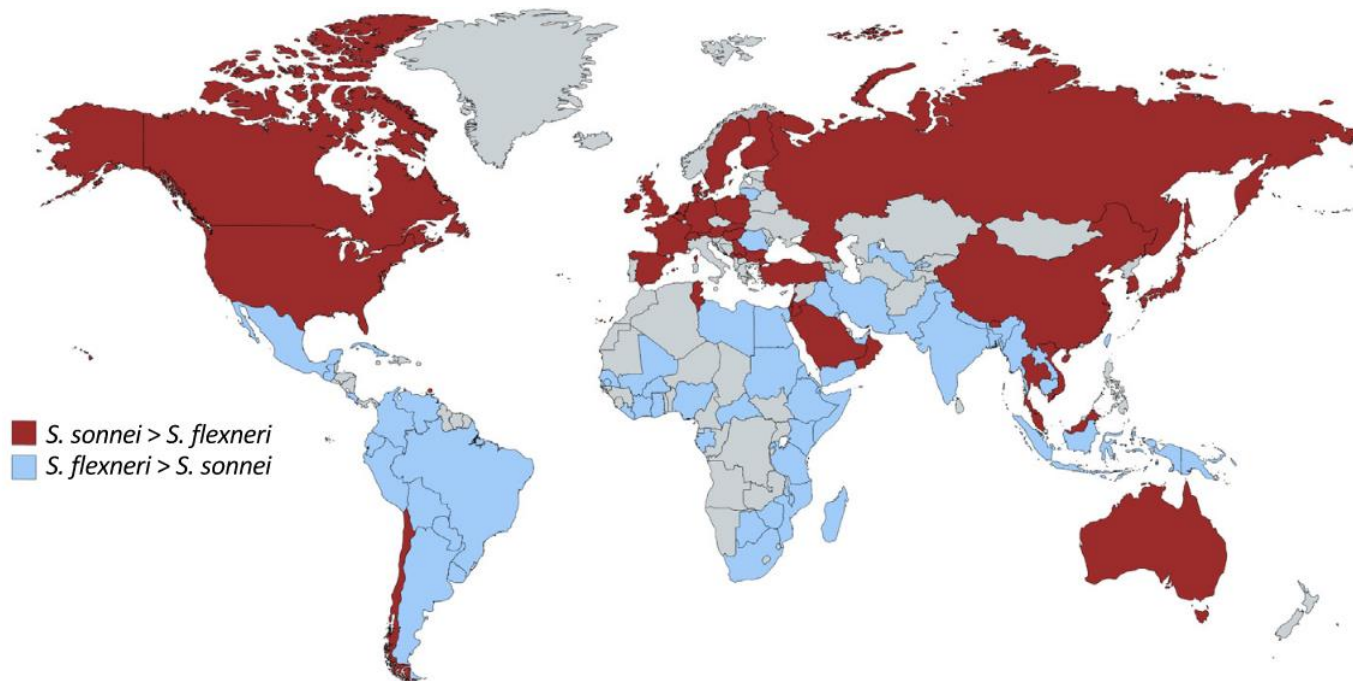
- Give an example of the use of WGS for national surveillance of bacterial pathogens
- Illustrate bioinformatic tools for analysis of WGS data:
 - Detection of **AMR** markers
 - Assignment of hierarchical **genotypes**
 - Visualization and annotation of **phylogenetic trees**
- Discuss advantages and limitations of WGS for surveillance

Shigella species and epidemiology

- *Shigella* are Gram-negative pathogenic enterobacteria
- Four species: *S. flexneri*, *S. sonnei*, *S. dysenteriae*, and *S. boydii*



Source: CDC, USA



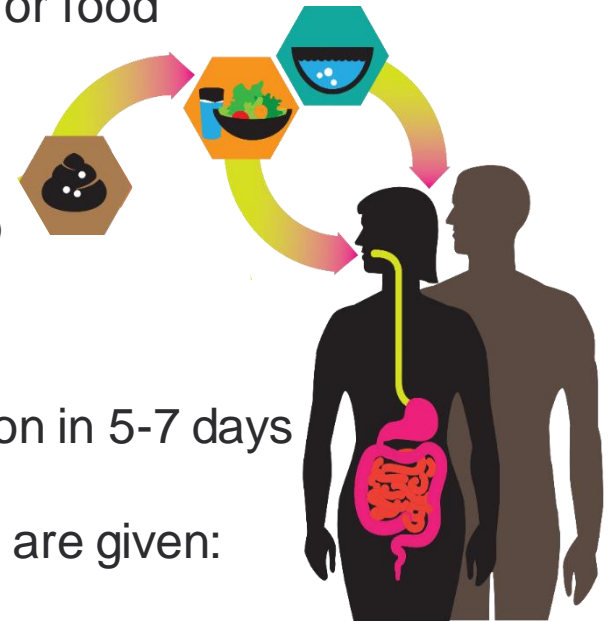
Shigella transmission and disease

Fecal-Oral transmission through contaminated water or food

→ Causing severe dysentery (diarrhea, fever, and stomach cramps)

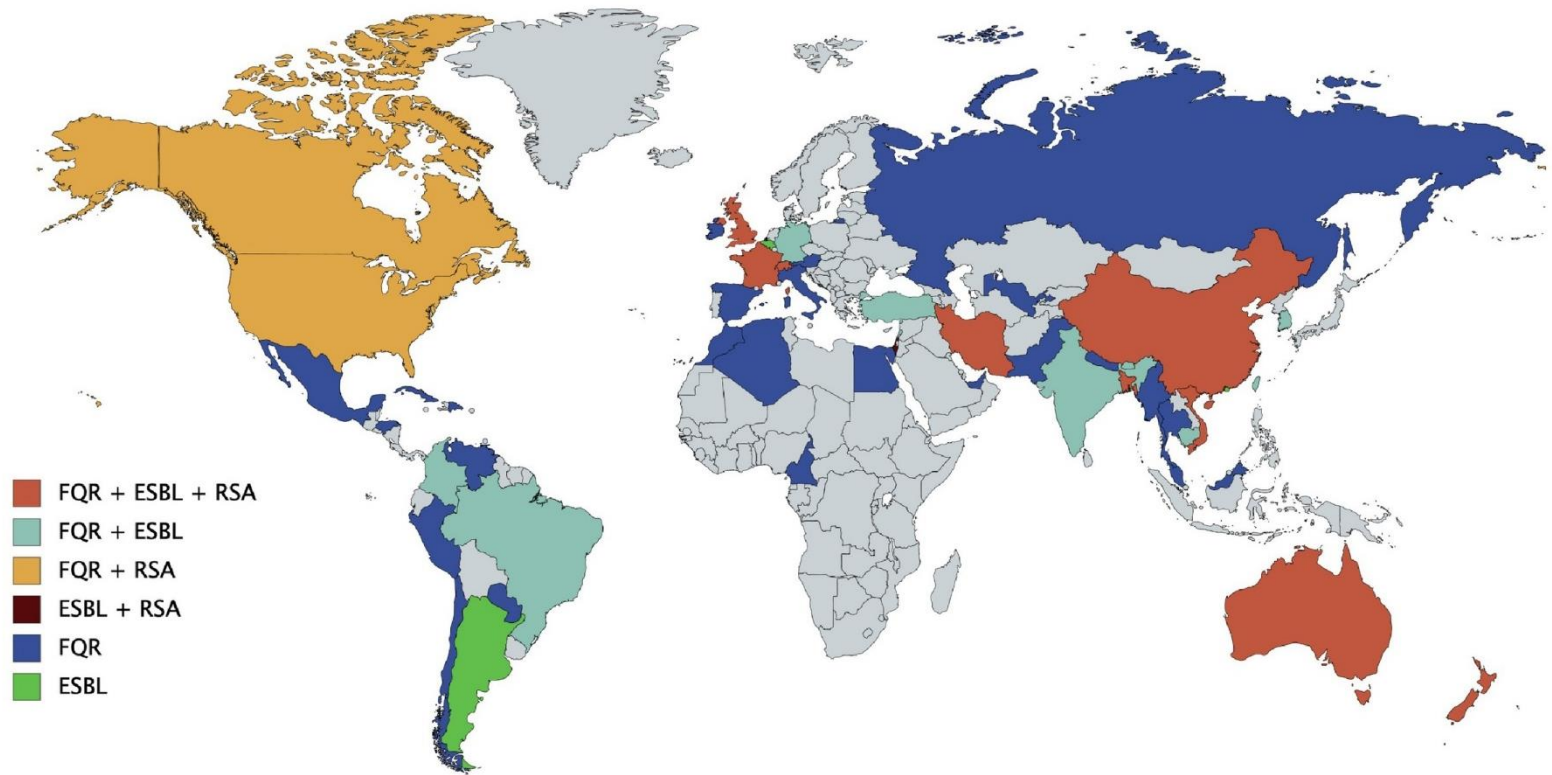


- Most cases resolve without medication in 5-7 days
- For severe cases first-line antibiotics are given:
 - Ciprofloxacin (Cip, adults)
 - Azithromycin (Azm, children)



www.waterpathogens.org

Global emergence of resistant *S. sonnei* strains



Trends in Microbiology

ESBL = extended-spectrum β -lactamase-producing (i.e., resistant to third-generation cephalosporins or carbapenem)
FQR = fluoroquinolone resistance (incl. Cip)
RSA = reduced sensitivity to azithromycin

AMR in *Shigella* is associated with men who have sex with men (MSM)

- England (Baker *et al.*, 2015, Mitchell *et al.*, 2019, Bardsley *et al.*, 2020)
- Taiwan (Chiou *et al.*, 2016)
- Canada (Gaudreau *et al.*, 2011, Yousfi *et al.*, 2019)
- Switzerland (Hinic *et al.*, 2018)
- Australia (Ingle *et al.*, 2019 & 2020)

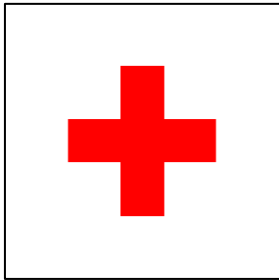


CDC info sheet on Shigellosis among MSM

Surveillance of *Shigellosis* in Belgium

~400 *Shigellosis* cases per year in Belgium 2013–2018 (Ventola *et al.*, 2019)

➤ 72 % caused by *Shigella sonnei* (*S. sonnei*)



Clinical laboratories



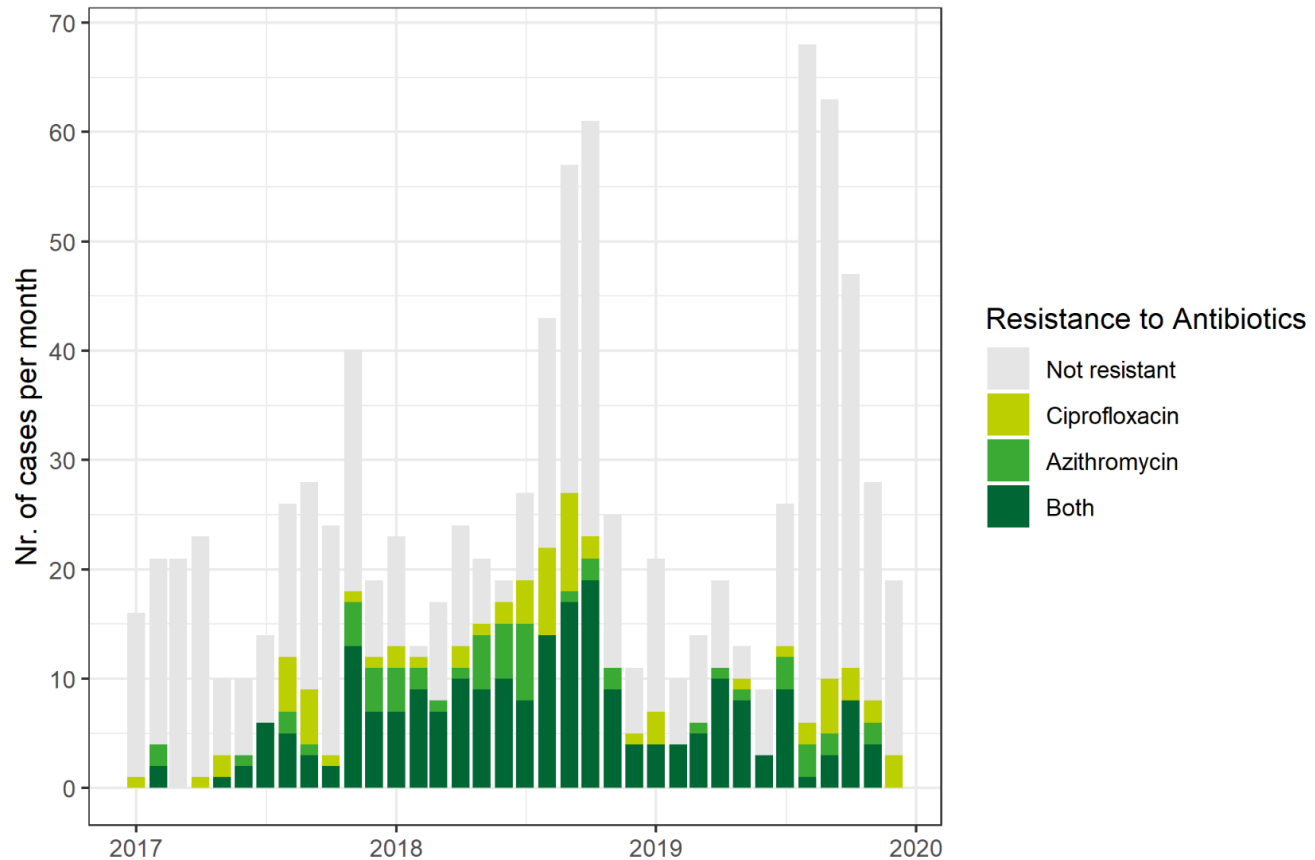
submit isolates and
patient information
(voluntary)

National Reference Centre for *Salmonella* and *Shigella* at Sciensano

- Typing of isolated strains to identify species and subtypes
- Determination of antibiotic resistance by broth microdilution

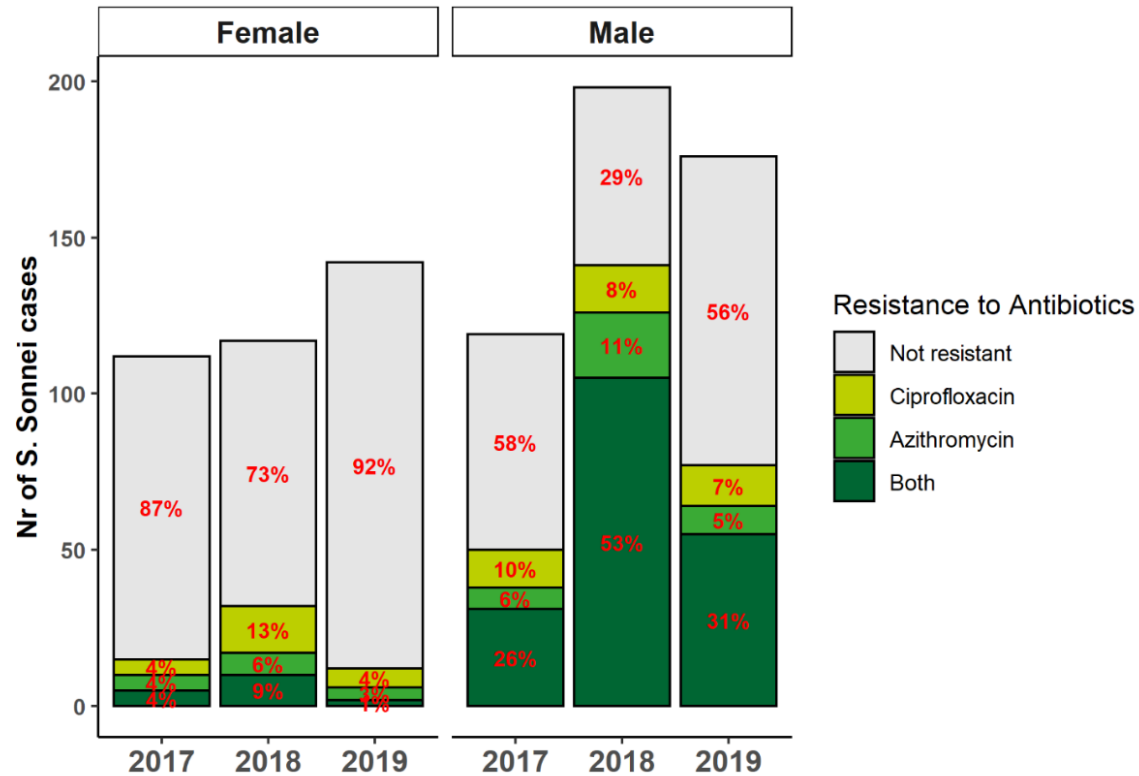


Epidemiology of *Shigellosis* in Belgium



37% of Belgian isolates were resistant to first-line antibiotics

Epidemiology of *Shigellosis* in Belgium



Men have an increased risk to carry a strain with resistance as compared to women

Why incorporate WGS in *Shigella* surveillance in Belgium?

- Difficulty to **differentiate and track** strains for public health surveillance or outbreak investigations using classical typing methods due to the limited genomic diversity of *S. sonnei*
- **Detection of AMR** in Belgium is currently limited to laboratory diagnostics
 - Testing for reduced sensitivity to Azithromycin (RSA) was only implemented in Belgium in 2017
- Difficulty to know the **origins or identify import events** of a strain
 - Travel history is only available for 10 % of cases in Belgium

Objective

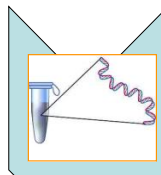
To describe clusters of multi-drug resistant *S. sonnei* in Belgium, through the use of whole genome sequencing (WGS), and to identify associated risk groups.

Methods

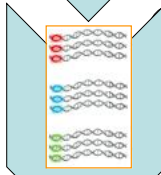
Sample selection:

- 372 Belgian *S. sonnei* isolates from 2013-2019
- 192 *S. sonnei* isolates from public databases (Latin-America, Asia, Europe, MSM-association)

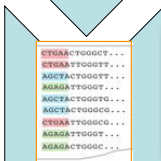
Sample processing:



Shigella isolates were cultured overnight in BHI broth (BD) at 37°C
DNA was extracted using MgC Bacterial DNA Kit™



Library prep was performed with Nextera XT DNA sample preparation kit



Sequencing of 250-bp paired-end reads on Illumina MiSeq



Using Qiagen CLC Genomic Workbench 20.0.2:
Reads were mapped to *S. sonnei* Ss046 reference genome
Single-nucleotide variants were identified
Neighbor Joining trees were constructed

Methods

Bioinformatics Platform: <https://usegalaxy.org/>

Galaxy Workflow Visualize Shared Data Help Login or Register Using 0%

Tools

search tools

Upload Data

Get Data

Collection Operations

GENERAL TEXT TOOLS

Text Manipulation

Filter and Sort

Join, Subtract and Group

Datamash

GENOMIC FILE MANIPULATION

FASTA/FASTQ

FASTQ Quality Control

SAM/BAM

BED

VCF/BCF

Nanopore

Convert Formats

Galaxy is an open source, web-based platform for data intensive biomedical research. If you are new to Galaxy start here or consult our help resources. You can install your own Galaxy by following the tutorial and choose from thousands of tools from the Tool Shed.

James P. Taylor Foundation for Open Science.

“The most important job of senior faculty is to mentor junior faculty and students.” — @jtxt

Announcing the James P. Taylor (JXTX) Foundation for Open Science

History

search datasets

Unnamed history

(empty)

This history is empty. You can load your own data or get data from an external source



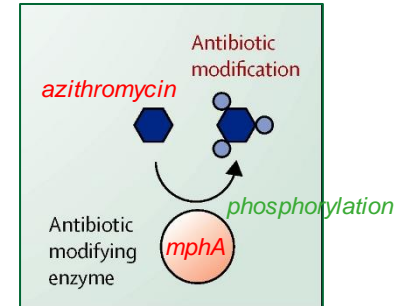
The Galaxy Team is a part of the Center for Comparative Genomics and Bioinformatics at Penn State, the Department of Biology at Johns Hopkins University and the Computational Biology Program at Oregon Health & Science University.

This instance of Galaxy is utilizing infrastructure generously provided by CyVerse at the Texas Advanced Computing Center, with support from the National Science Foundation.

Methods: Bioinformatic detection of AMR markers

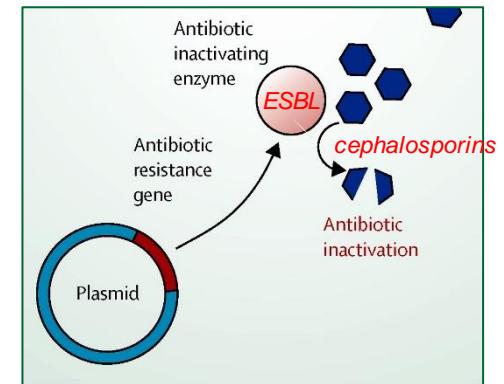
ResFinder: presence of **AMR genes** (Bortolaia *et al.* 2020)

- macrolide resistance genes (*mphA*, *ermB*)
- resistance to azithromycin



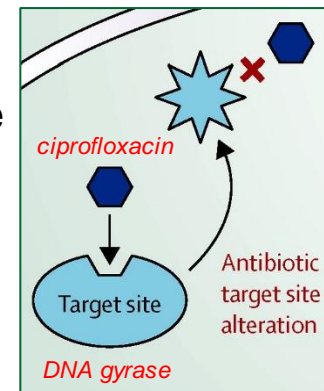
PlasmidFinder: detection of **plasmid replicons** (Carattoli *et al.* 2014)

- Inc type plasmids
- carry extended-spectrum-beta lactamases (ESBL genes)
- resistance to third generation cephalosporins



PointFinder: detection of **chromosomal mutations** (Zankari *et al.* 2017)

- quinolone resistance determining region (QRDR) of bacterial DNA gyrase
- *gyrA* (S83L), *gyrA* (D87G), *gyrA* (D87Y) and *parC* (S80I)
- resistance to ciprofloxacin



Methods: Assignment of hierarchical genotypes

MDR.fastq.gz.json - Mykrobe

WORKS OFFLINE

MYKROBE

ALL DRUGS EVIDENCE SPECIES SAVE NEW

FIRST LINE DRUGS

- Isoniazid **RESISTANT**
- Rifampicin **RESISTANT**
- Ethambutol **SUSCEPTIBLE**
- Pyrazinamide **SUSCEPTIBLE**

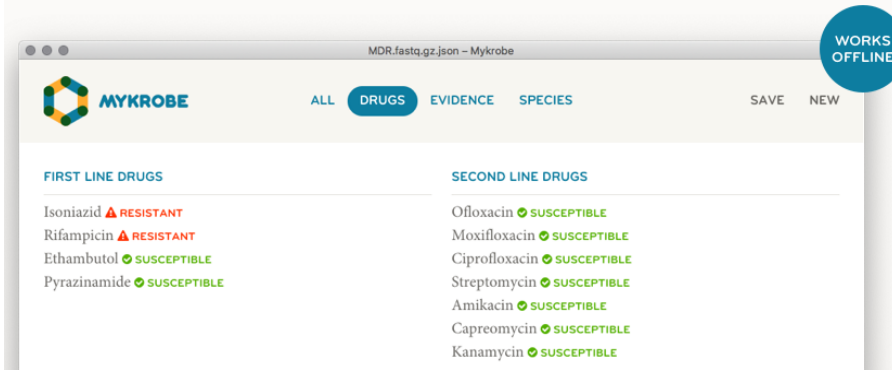
SECOND LINE DRUGS

- Ofloxacin **SUSCEPTIBLE**
- Moxifloxacin **SUSCEPTIBLE**
- Ciprofloxacin **SUSCEPTIBLE**
- Streptomycin **SUSCEPTIBLE**
- Amikacin **SUSCEPTIBLE**
- Capreomycin **SUSCEPTIBLE**
- Kanamycin **SUSCEPTIBLE**

Hunt M, Bradley P, Lapierre SG et al.
Antibiotic resistance prediction for
Mycobacterium tuberculosis from genome
sequence data with Mykrobe.
Wellcome Open Res 2019

→ <https://www.mykrobe.com/>

Methods: Assignment of hierarchical genotypes



Hunt M, Bradley P, Lapierre SG et al. Antibiotic resistance prediction for *Mycobacterium tuberculosis* from genome sequence data with Mykrobe. Wellcome Open Res 2019

→ <https://www.mykrobe.com/>

Integrated hierarchical SNV-based genotyping scheme for *S. sonnei* (Hawkey et al. 2021)

→ <https://github.com/katholt/sonneityping>

New genotyping framework		Previously defined as		Description
Genotype	Name	Name	Study	
Lineage II				
2.8	Korea II	Korea II	Holt 2012	Associated with Korea
2.9, 2.10, 2.11	Latin America II	South America II LA sublineage IIa & IIb	Holt 2012 Baker 2017	Associated with Latin America
Lineage III				
3.4	Latin America III	South America III LA sublineage IIIa & IIIb	Holt 2012 Baker 2017	Associated with Latin America
3.6	Central Asia III	Central Asia IIIa	Holt 2012	Associated with Central Asia
3.6.1	CipR parent	-	This study	Subclade from which ciprofloxacin-resistant sublineage emerged
3.6.1.1	CipR	Ciprofloxacin-resistant Pop2	The 2015 The 2019	Ciprofloxacin-resistant triple mutation sublineage

Methods: Visualization and annotation of the phylogenetic tree

ggtree: visualization and annotation of phylogenetic trees

release version 1.14.6 devel version 1.15.5 download 42242/total download 1656/month

The `ggtree` package extending the `ggplot2` package. It based on grammar of graphics and takes all the good parts of `ggplot2`. `ggtree` is designed for not only viewing phylogenetic tree but also displaying annotation data on the tree. `ggtree` is released within the [Bioconductor](#) project and the source code is hosted on [GitHub](#).

Authors

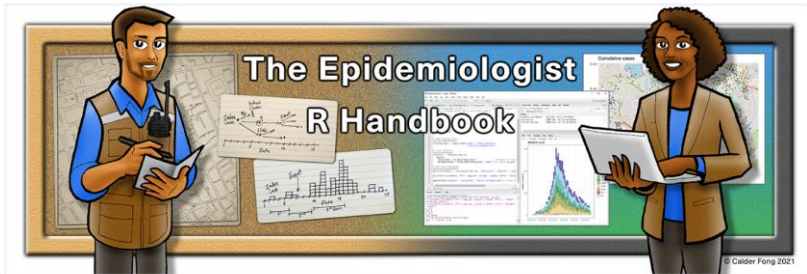
Guangchuang Yu, School of Basic Medical Sciences, Southern Medical University.

<https://yulab-smu.top/treedata-book/>

<https://github.com/YuLab-SMU/ggtree>



Resources and R Code for phylogenetic analysis



<https://epirhandbook.com/>

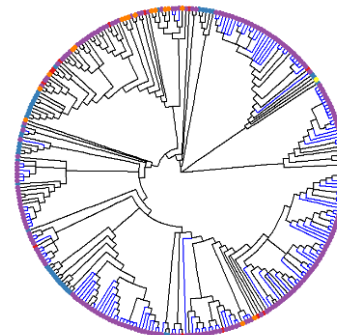
38 Phylogenetic trees

38.1 Overview

Phylogenetic trees are used to visualize and describe the relatedness and evolution of organisms based on the sequence of their genetic code.

They can be constructed from genetic sequences using distance-based methods (such as neighbor-joining method) or character-based methods (such as maximum likelihood and Bayesian Markov Chain Monte Carlo method). Next-generation sequencing (NGS) has become more affordable and is becoming more widely used in public health to describe pathogens causing infectious diseases. Portable sequencing devices decrease the turn around time and hold promises to make data available for the support of outbreak investigation in real-time. NGS data can be used to identify the origin or source of an outbreak strain and its propagation, as well as determine presence of antimicrobial resistance genes. To visualize the genetic relatedness between samples a phylogenetic tree is constructed.

In this page we will learn how to use the **ggtree** package, which allows for combined visualization of phylogenetic trees with additional sample data in form of a dataframe. This will enable us to observe patterns and improve understanding of the outbreak dynamic.

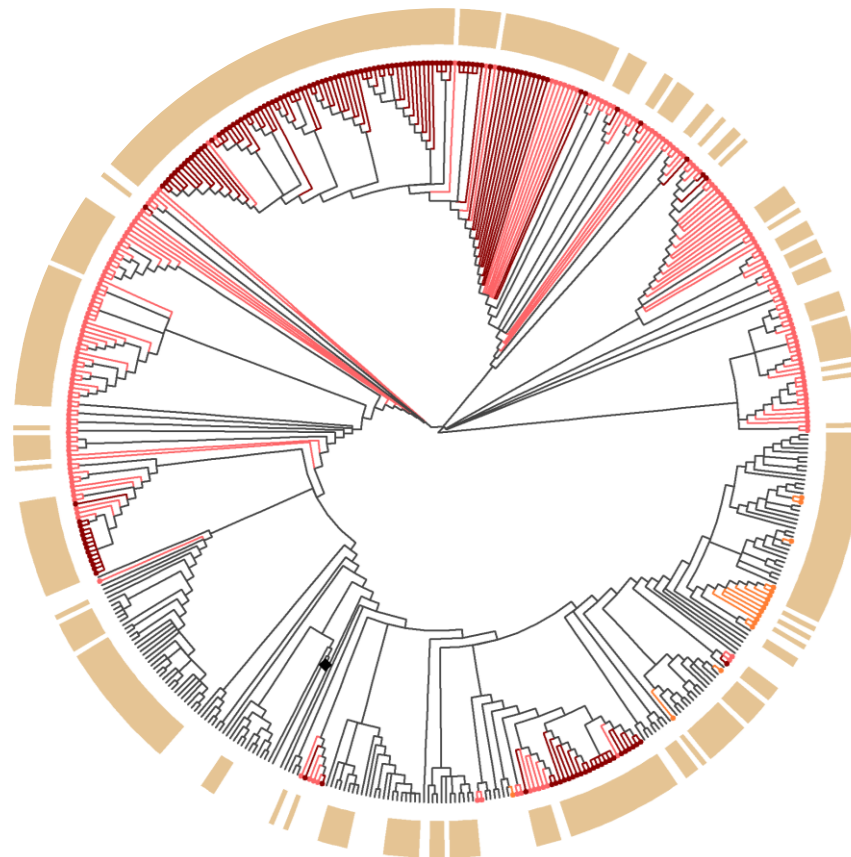


Sample Origin — NRCSS Belgium — Other

Continent

• Africa • Australia • Latin America
• Asia • Europe • North America

Results: Phylogenetic analysis reveals several distinct clusters with double resistance to first-line antibiotics

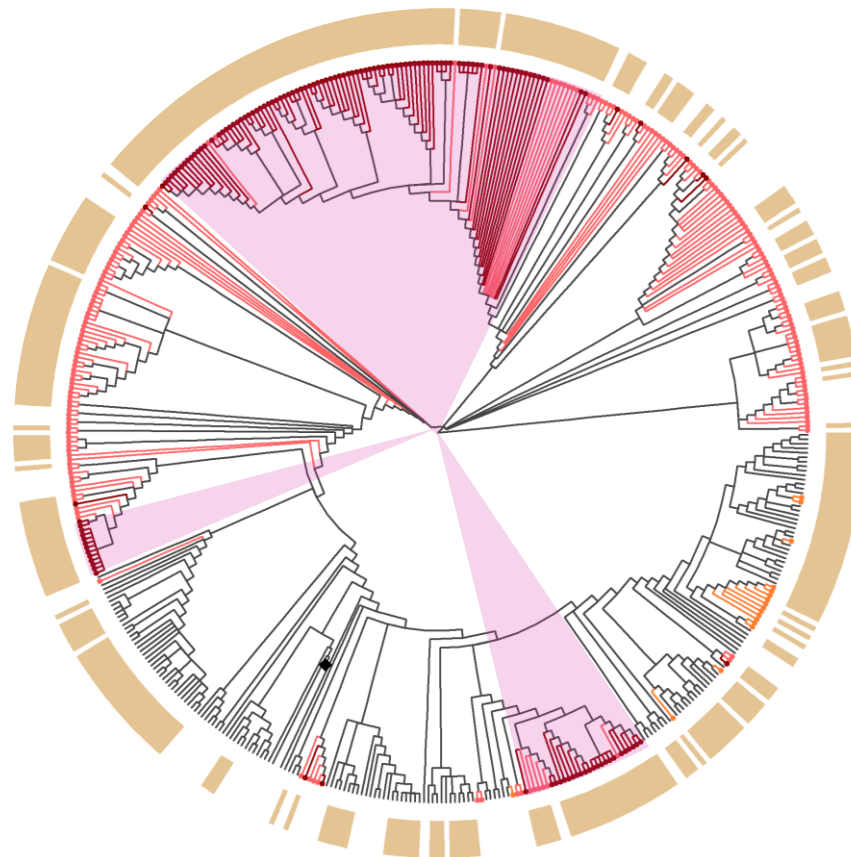


Antibiotic resistance

- Both
- Azithromycin
- Ciprofloxacin

NRC Belgium Yes

Results: Phylogenetic analysis reveals several distinct clusters with double resistance to first-line antibiotics

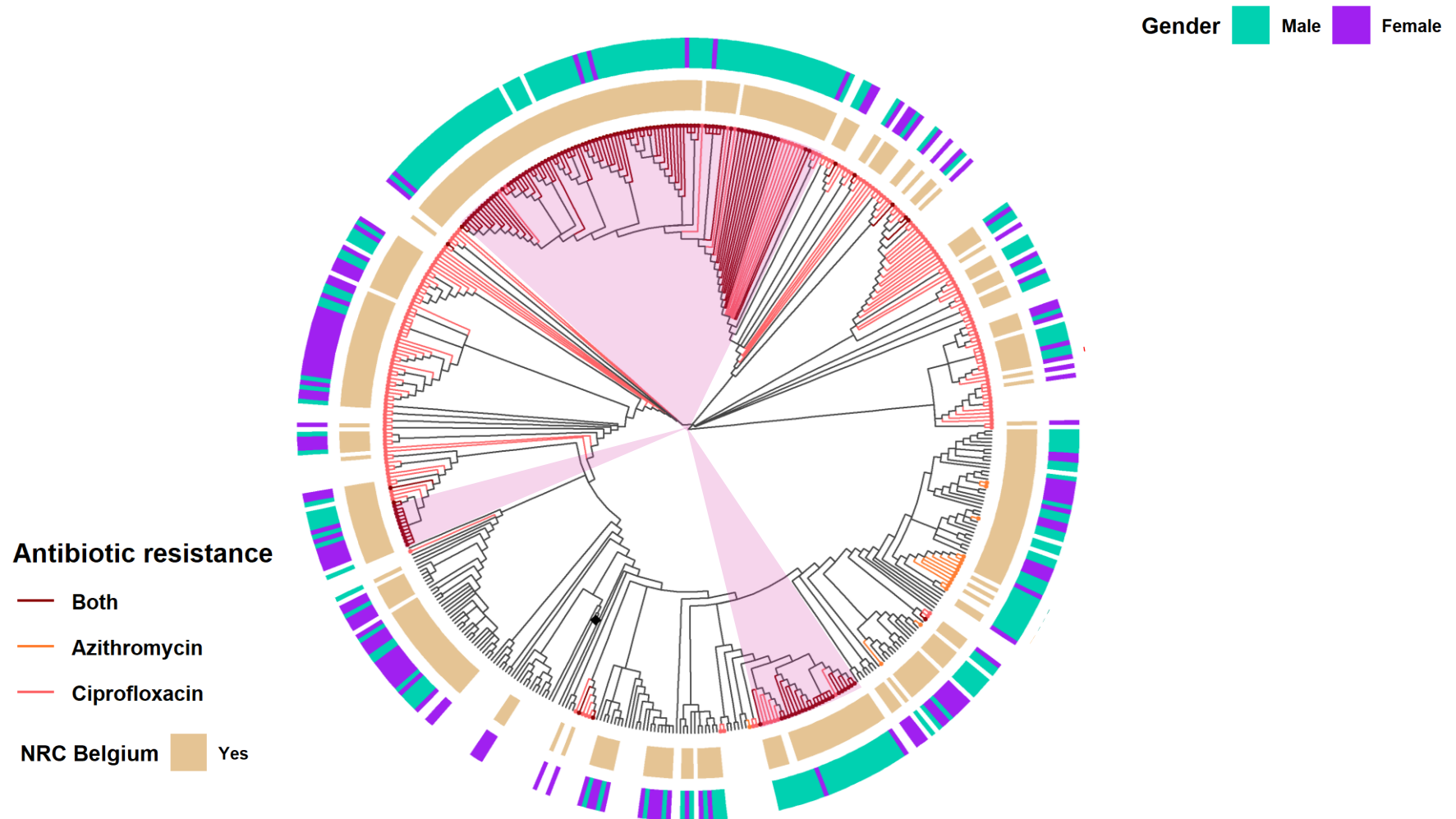


Antibiotic resistance

- Both
- Azithromycin
- Ciprofloxacin

NRC Belgium Yes

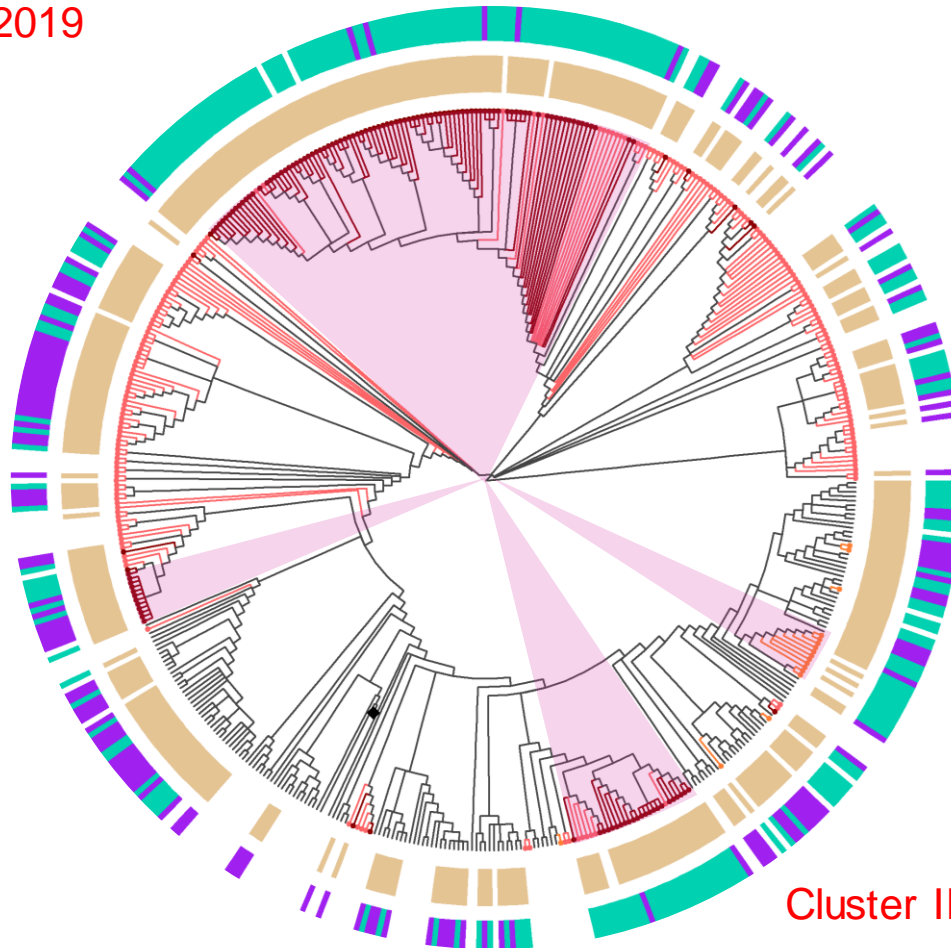
Results: Resistant clusters are predominantly comprised of male cases



Results: Resistant clusters are predominantly comprised of male cases

Cluster I: 102 cases, 94% ♂
2015-2019

Gender ■ Male ■ Female



Antibiotic resistance

- Both
- Azithromycin
- Ciprofloxacin

NRC Belgium ■ Yes

Cluster III: 12 cases, 92% ♂
2013-2018

Cluster II: 21 cases, 95% ♂
2015-2019

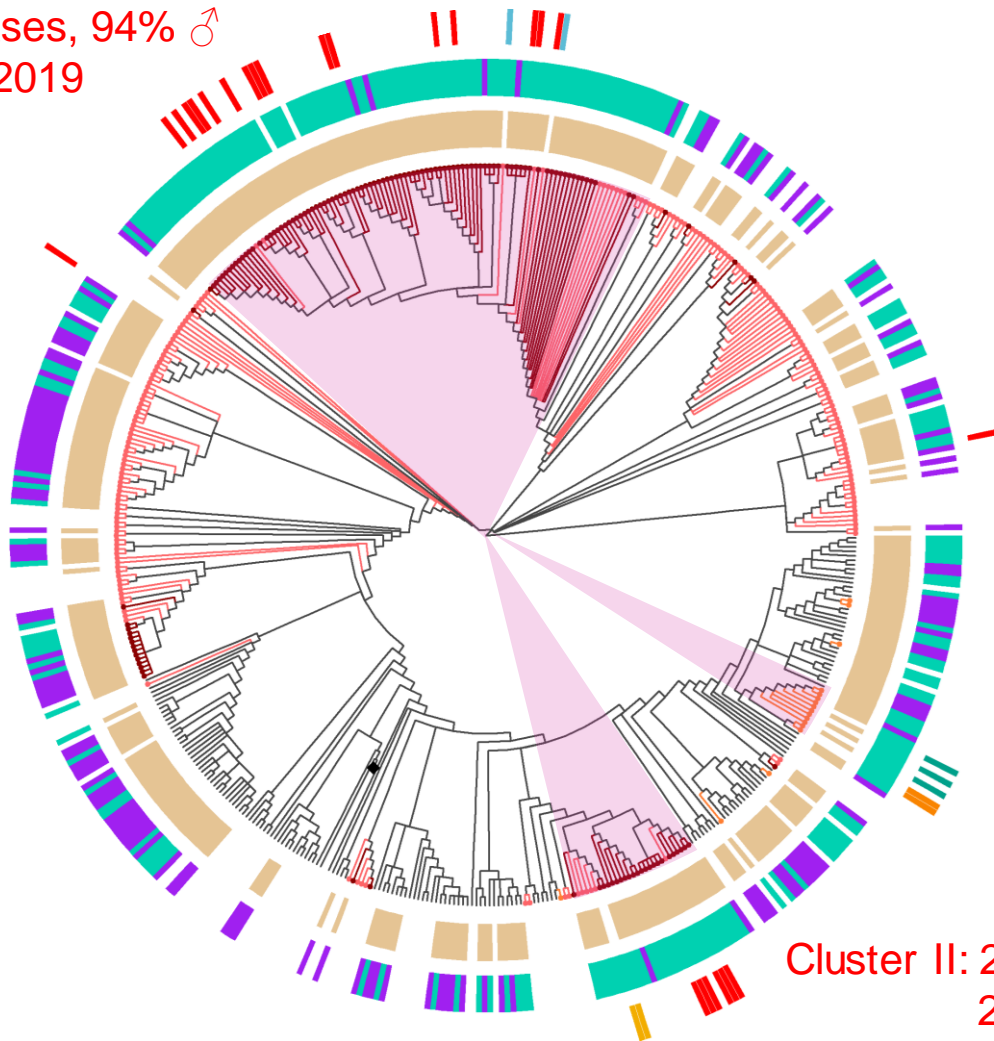
Results: Resistant clusters are associated with men who have sex with men (MSM)

Cluster I: 102 cases, 94% ♂
2015-2019

Gender ■ Male ■ Female

MSM association:

- MSM Belgium
- MSM UK Clade 1
- MSM UK Clade 2
- MSM UK Clade 3
- MSM UK Clade 7



Antibiotic resistance

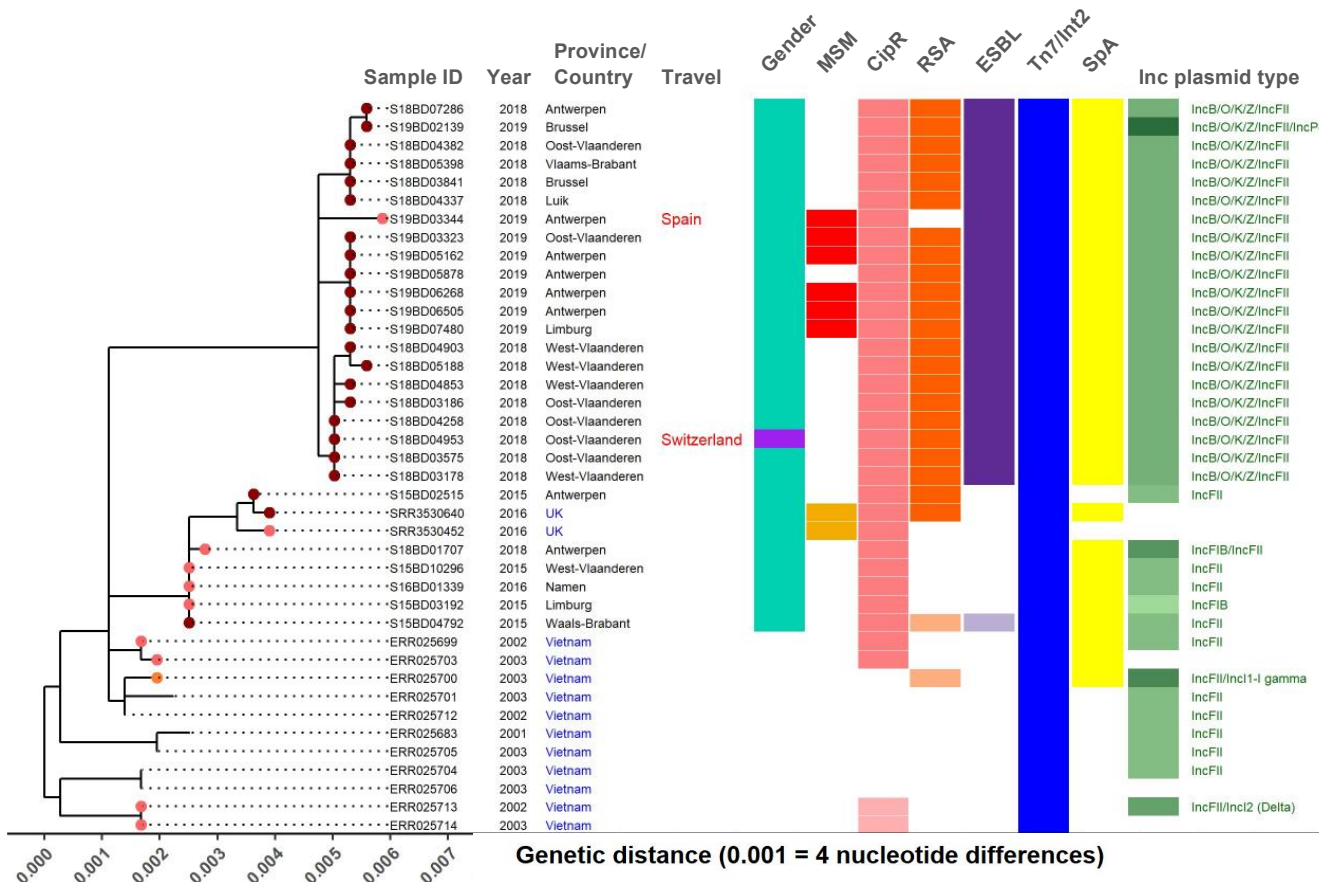
- Both
- Azithromycin
- Ciprofloxacin

NRC Belgium ■ Yes

Cluster III: 12 cases, 92% ♂
2013-2018

Cluster II: 21 cases, 95% ♂
2015-2019

Results: Detailed analysis of Cluster II revealed evolution of multi-drug resistance from Vietnamese strains



Antibiotic resistance ● RSA & CipR ● RSA ● CipR

Gender ■ Male ■ Female

.....S15BD0161 2015 LI
S17BD03549 2017 AI

AMR markers:

QRDR mutations ■ *gyrA* (S83L) ■ *gyrA* (D87G)

Macrolide resistance genes ■ *mph(A)* ■ *mph(A)* & *erm(B)*

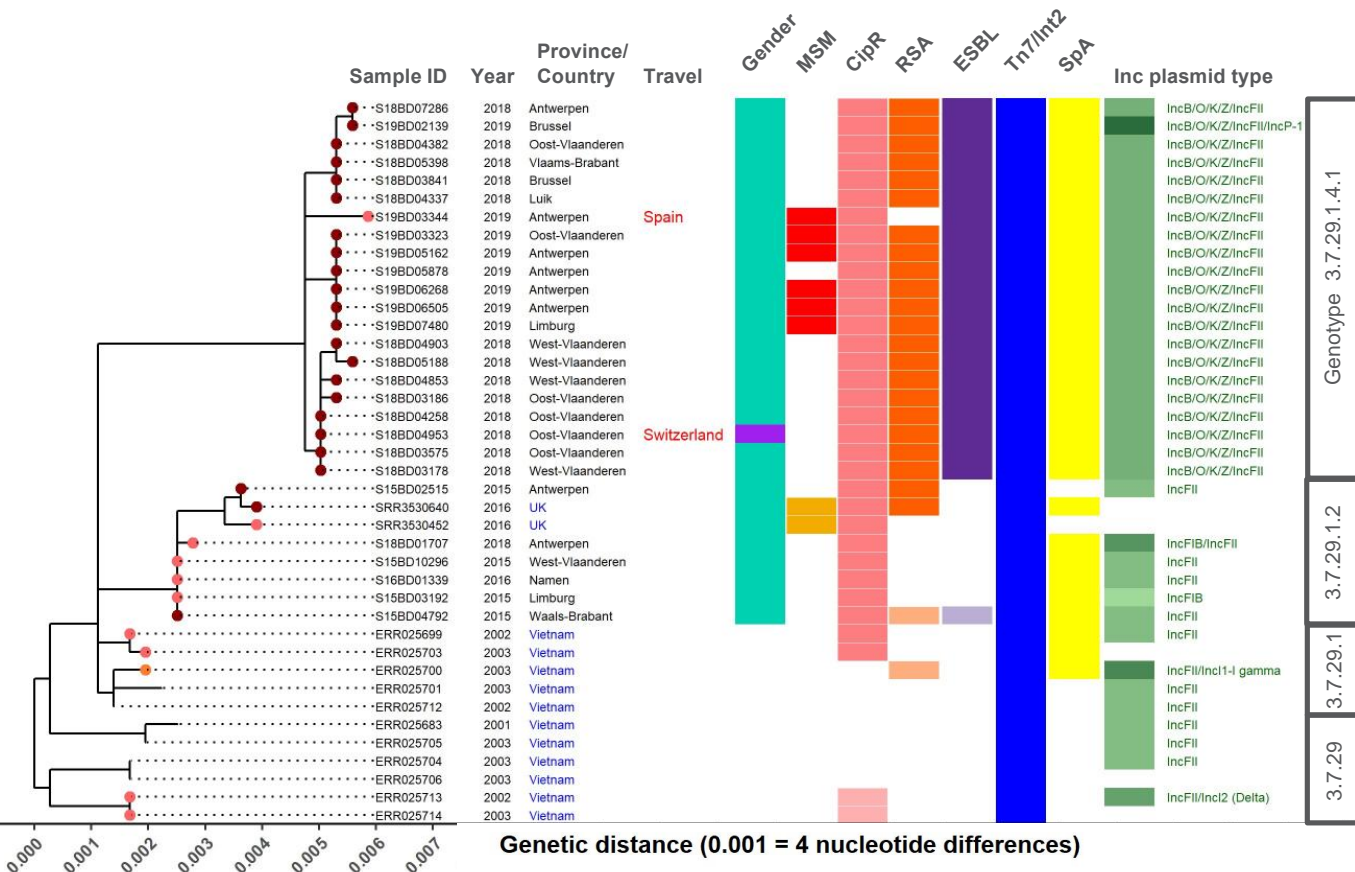
Chromosomal resistance island ■ Tn7/Int2

ESBL genes ■ *blaCTX-M-27* ■ *blaDHA-1*

Plasmids ■ SpA

■ - ■ Other Inc type plasmids

Results: Genotype assignment with Mykrobe revealed two internationally circulating MSM-related clusters in Belgium



Outbreaks in MSM in Australia 2016-2019 (Ingle *et al.*, 2019 & 2020)

Circulation among MSM in the UK (Bardsley *et al.* 2020)

Antibiotic resistance ● RSA & CipR ● RSA ● CipR

Gender ■ Male ■ Female

.....S15BD07161 2015 LI

.....S17BD03549 2017 AI

MSM UK Clade 2

AMR markers:

QRDR mutations ■ *gyrA* (S83L) ■ *gyrA* (D87G)

Macrolide resistance genes ■ *mph(A)* ■ *mph(A)* & *erm(B)*

Chromosomal resistance island ■ Tn7/Int2

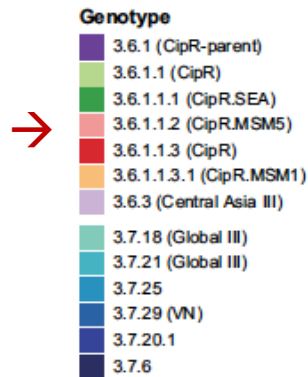
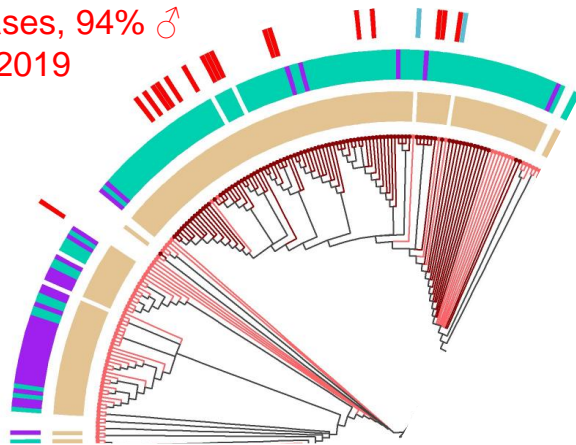
ESBL genes ■ *bla*CTX-M-27 ■ *bla*DHA-1

Plasmids ■ SpA

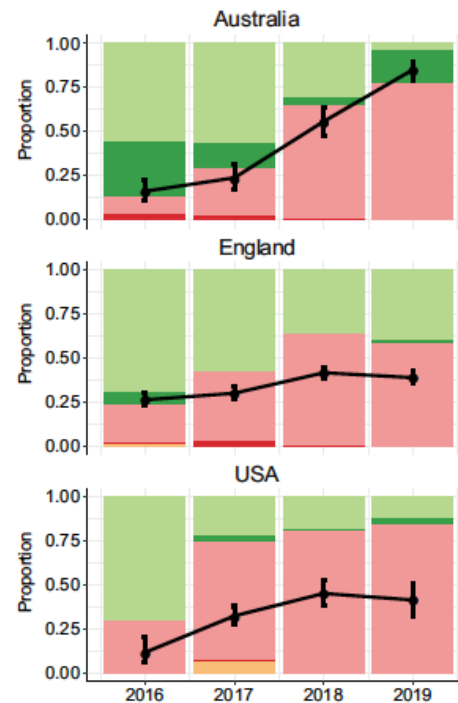
■ - ■ Other Inc type plasmids

Belgian cluster 1 was of genotype 3.6.1.1.2, linked to increased resistance in ciprofloxacin and azithromycin in Australia, England and the USA

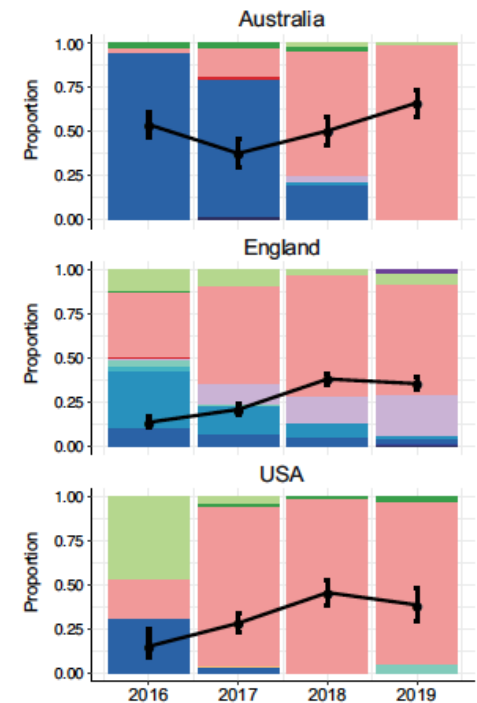
Cluster I: 102 cases, 94% ♂
2015-2019



a Ciprofloxacin resistance



b Azithromycin resistance



Conclusions

- More than **1/3** of all *S. sonnei* strains isolated in Belgium 2017-2019 harbor resistance to at least one important first-line antibiotic
- Men have **a higher risk** to carry strains with antibiotic resistance
- WGS identified several clusters of **multi-drug resistant** *S. sonnei* strains, which are continuously circulating in the **MSM-community** in Belgium and are of the same genotype as MSM-related isolates from other parts of the world

Recommendations

Increased Surveillance:

- Reporting and strain collection across all three regions in Belgium
- Inclusion of whole genome sequencing in routine surveillance
- Enhanced contact tracing of multi-resistant cases of *Shigellosis*

Case Management:

- Include AMR testing and improve antimicrobial stewardship

Assessment of the epidemiological situation in MSM in Belgium:

- Data collection on sexual orientation
- Further case control studies

Prevention and education campaigns:

- Raising awareness of *Shigellosis* as a sexually transmittable infection

WGS for surveillance can be useful to:

- Monitor the circulation of strains on a national as well as international level
- Detect a variety of AMR markers – independent of laboratory constraints
- Generate hypothesis about origins of strains and history of acquisition of AMR

Open-source tools, code and data sharing are needed to bridge current gaps in surveillance and build global capacity for WGS and data analysis

Limitations and considerations for WGS

- Proper sample collection, transport and processing is necessary to obtain sufficient genetic material of high quality
- Unique identifiers and proper labelling are needed to link samples to patient data
- Voluntary isolate submission may introduce bias due to non-exhaustive sample collection
- Missing isolates and data from countries without capacity for WGS may introduce bias in global dissemination of strains

THANK YOU FOR YOUR ATTENTION

Sciensano:

Pieter-Jan Ceysens
An van den Bossche
Wesley Mattheus
Margo Maex
Dieter van Cauteren

AZG:

Valeska Laisnez
Naïma Hammami

Fellowship Funding:

ECDC

